SOP: **Propagation of Prostate Epithelial Cells (PrEC, Lonza Biosciences)**
Date modified: **10/6/2010**
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Ordering Information

Prostate Epithelial Cells (PrEC) may be ordered either as frozen ampoules or as starter cultures. The former contain ~0.5-1 x 10^6 cells; the latter are initiated at Lonza and sent in a T225 flask containing ~6-7 x 10^6 cells.

To order frozen ampoules + media:

Name:	PrEC – Prostate Epithelial Cell
Item #:	CC-2555 (PrEC - Cryopreserved ampoule)
	CC-3166 (PrEGM™ BulletKit® = CC-3165 + CC-4177)

To order starter cultures:

Name:	PrEC – Prostate Epithelial Cell
Item #:	CC2555T225 (PrEC in PrEGM™ T225 Flask)
	CC-3166 (PrEGM™ BulletKit® = CC-3165 + CC-4177)

Notes:

The number of BulletKits purchased depends on the target number of cells to be generated. A rule of thumb is 10 BulletKits for every initial T225 flask of cells. It is strongly recommended to purchase all of the media that will be required for a complete expansion series, since media supply may be erratic.

Materials List

1. Cell-type specific medium (BulletKits – Lonza Biosciences)
2. T225 tissue culture flasks
3. Corning conical centrifuge tubes (15mL and 50mL)
4. Graduated pipets (1, 5, 10, 25, 50mL)
5. Pen-Strep solution (if required; Lonza typically supplies antibiotics)
6. Phosphate Buffered Saline (1X PBS) (Cellgro, Cat# 21-040-CM)
7. Accutase Enzyme Cell Detachment Medium (EBiosciences Cat# 00-4555)
8. Eppendorf Centrifuge 5810R
9. Hemocytometer
10. Micropipet w/ P20 tips
11. Microscope

Procedure

A. Receipt of proliferating cells

1) Swab down flask with 70% ethanol.
2) Equilibrate for 3-4 hours in 37°C, 5% CO_2 humidified incubator.
3) Remove shipping medium. Replace with fresh medium and return to incubator.

B. Sub-culture

1) Propagate cells until density reaches 70-90% confluence.
2) Aspirate medium.
3) Wash cells with warm 1X PBS.
4) Add 15mL of Accutase and return to incubator for 10-15 minutes, or until cells detach.
5) Immediately remove cells, rinse flask with warm 1X PBS to collect residual cells, and pellet at 500 x g for 5 minutes (4^oC).
6) Gently re-suspend cell pellet in warm medium.
7) Count cells with hemocytometer.
8) Add warmed medium to flasks.
9) Seed flasks at **2,500 cells/cm^2 density.**
10) Record each subculture event as a passage.

C. Maintenance

1) Change media the day after seeding and every OTHER day thereafter.
2) Increase media volume as confluency increases (volumes assume the use of
3) T225 flasks):
    a. 25% = 1mL/5 cm^2
    b. 25-45% = 1.5mL/5 cm^2
    c. 45%+ = 2mL/5 cm^2.
4) Per the above an exemplary schedule might be:
    a. day 1, plate into T225: use 50mL of media.
    b. day 2, change media, use 50mL of media.
    c. day 4, change media, use 100mL of media (if confluency is >50%).
    d. day 6, change media, use 100mL of media (or harvest if ready).
    e. day 7 or 8 (harvest when cells reach 6 x 10^6 cells/flask).

D. Harvest

1) Pass cells 3-4 times until the desired cell number is achieved (primary cells will senesce after 4-5 passages).
2) Remove cells from flasks according to protocol described above under “Sub-culture.”
3) Examine viability using Trypan blue staining (SOP TP-7).