SOP: Propagation of Renal Cell Carcinoma (RCC) RCC_7860
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Ordering Information

RCC_7860 cells can be ordered from ATCC as a frozen ampule (Cat# CRL-1932).

Name: 786-0, Renal Cell Adenocarcinoma

Sex: Male

Notes:
This is an adherent cell line derived from a primary renal cell adenocarcinoma. The cells display both microvilli and desmosomes, and can be grown in soft agar. The cells produce a PTH like peptides that is identical to peptides produced by breast and lung tumors. The peptide has an N terminal sequence similar to PTH, has PTH like activity, and has a molecular weight of 6000 daltons. Cytogenetic analysis indicates that the cells are hypertriploid.

Materials List

1. DMEM, High Glucose, Pyruvate (Cat# 11995 Gibco)
2. Fetal Bovine Serum (Cat#100-106, Gemini Bio-Products)
3. Non-Essential Amino Acids (Cat#11140 Gibco)
4. L-glutamine (Cat#25030 Gibco)
5. 0.5% Trypsin/0.1%EDTA (Cat# 25300 Gibco)
6. 10cm culture plates
7. Graduated pipets (1, 5, 25mL)
8. Hemocytometer
9. Microscope

Growth Medium for 786-0

DMEM, High Glucose, Pyruvate
10% Fetal Bovine Serum
1% Non-Essential Amino Acids
1% L-glutamine

Procedure

A. Receipt of frozen cells and starting cell cultures.

1) Immediately place frozen cells in liquid nitrogen storage incubator.
2) Quickly thaw ampoule in 37^0C water bath
3) Transfer thawed cells to a 10cm plate with 10mL of warm growth media.
4) Allow cells to recover over night in 37^0C, 5% CO_2 humidified incubator.
5) Pour off medium the next day, replace with fresh medium and return to incubator.

B. Sub-culture

1) Propagate cells until density reaches 80-90% confluence.
2) Decant medium.
3) Wash cells with warm 1X PBS.
4) Add 2 ml of Trypsin/EDTA and return to incubator for 5 minutes.
5) Add 6 ml of fresh medium and resuspend cells by gently pipetting.
6) Perform 1:3 to 1:10 cell split as needed.
7) Record each subculture event as a passage.

C. Maintenance

1) Change media the day after seeding and 1-2 times per week thereafter.

Use ~10 mLs of medium per 10cm plate.

D. Harvest

1) Remove cells from plates according to protocol described above under 'subculturing'
2) Examine viability using trypan blue staining (SOP)

E. Passaging

1) Recommend no more than 25 – 30 passages.